PROGESTERONE (PROG) CHEMILUMINESCENCE IMMUNOASSAY

KIT
Catalog No. CL1106-2

INTENDED USE
The Autobio progesterone CLIA test kit is intended for the quantitative determination of progesterone concentration in human serum.

INTRODUCTION
Progesterone (Pregn-4-ene-3,20-dione) is a C21 steroid hormone. The molecular weight of this steroid hormone is 314.5. Progesterone is a female sex hormone which, in conjunction with estrogens, regulates the accessory organs during the menstrual cycle and it is particularly important in preparing the endometrium for the implantation of the blastocyte and in maintaining pregnancy. In non-pregnant whereas in pregnancy the placenta becomes the major source. Minor sources are the adrenal cortex for both sexes and the testes for males.

Progesterone circulates in blood mainly bound to Corticosteroid Binding globulin (CBG), Sec Hormon Binding globulin (SHBG) and Albumin. Only 2-10% of the total concentration circulates as free hormone. The measurements of plasma progesterone are clinically used to confirm ovulation and normal function of the corpus luteum in non-pregnant women.

Abnormal progesterone secretion has been implicated in premenstrual tension, irregular shedding of endometrium, dysmenorrhoea, and luteal insufficiency.

PRINCIPLE OF THE TEST
In the Prog CLIA test procedure, the reference standards and patient serum specimens are incubated with the progesterone antibody and the progesterone-horseradish peroxidase conjugate in the anti-mouse IgG coated wells. In this solid-phase system, the antibody bound progesterone will remain on the well while unbound progesterone will be removed by washing. A chemiluminescence reaction is developed when the CLIA substrates are mixed with the antibody bound progesterone-horseradish peroxidase enzyme conjugate. The Related Light Unit (RLU) is proportional to the amount of enzyme present and is inversely related to the amount of unlabeled Prog in the sample. By reference to a series of Prog reference standards assayed in the same way, the concentration of Prog in the unknown sample is quantified.

MATERIALS PROVIDED
1. Antibody Coated Microtiter Plate: Microplate coated with goat anti-mouse IgG (1 plate, 48 wells/96wells)
2. Enzyme Conjugate Reagent: Horseradish peroxidase (HRP) labeled progesterone (Prog) in Stabilizing Buffer (1 vial, 3.0ml/6.0 ml)
3. Anti-Prog Reagent: Mouse monoclonal antibodies to progesterone (anti-Prog MAb) in Stabilizing Buffer (1 vial, 3.0ml/6.0 ml)
4. Reference Standards: 0, 0.5, 1, 2.5, 10 and 30ng/ml (6 vials, Lyophilized)
5. Substrate A: (1 vial, 1.8ml/3.5ml)
6. Substrate B: (1 vial, 1.8ml/3.5ml)
7. PBS-T Powder: PBS-Tween (1 bag, 5g)

MATERIALS NOT PROVIDED
The following materials are required but not provided in the kit.
1. Precision pipettes and tips, 0.025ml, 0.05ml, 0.1ml
2. Distilled water.
3. Vortex mixer
4. Magnetic stirrer
5. Absorbent paper or paper towel
6. Graph paper
7. Luminometer

STORAGE OF TEST KIT AND INSTRUMENTATION
1. Unopened test kits should be stored at 2 ~ 8°C upon receipt and the microtiter plate should be kept in a sealed bag with desiccants to minimize exposure to damp air. The test kit may be used throughout the expiration date of the kit. Refer to the package label for the expiration date.

2. Opened test kits will remain stable until the expiration date shown, provided it is stored as prescribed above.

**SPECIMEN COLLECTION AND PREPARATION**

1. Blood should be drawn using standard venipuncture techniques and the serum should be separated from the red blood cells as soon as practical. Avoid grossly hemolytic, lipemic or turbid samples.

2. Plasma samples collected in tubes containing EDTA, heparin, or oxalate may interfere with test procedures and should be avoided.

3. Specimens should be capped and may be stored up to 48 hours at 2 ~ 8°C, prior to assaying. Specimens held for a longer time can be frozen at -20°C. Thawed samples must be mixed prior to testing. Multiple freeze-thaw cycles should be avoided.

**PRECAUTIONS AND WARNINGS**

1. For in vitro diagnostic use only.

2. Handling of reagents, serum specimens should be in accordance with local safety procedures.

3. The reference standards contain human source components, which have been tested and found non-reactive for hepatitis B surface antigen as well as HIV antibody. All animal products and derivatives have been collected from healthy animals. Bovine components originate from countries where BSE has not been reported. Nevertheless, the reference standards and components containing animal substances should be treated as potentially infectious.

4. Avoid any skin contact with all reagents.

5. Do not smoke, drink, eat or apply cosmetics in the working area. Do not pipette by mouth. Use protective clothing and disposable gloves.

**REAGENT PREPARATION**

1. All reagents should be brought to room temperature (18 ~ 25°C) before use. All reagents should be mixed by gently inverting or swirling prior to use. Do not induce foaming.

2. Reconstitute each lyophilized standard with 0.5 ml distilled water. Allow the reconstituted material to stand for at least 10 minutes. Reconstituted standards should be stored sealed at 2 ~ 8°C. Reconstituted standards should be used within 14 days and be frozen at -20°C for long term storage.

3. Prepare Wash Solution: add 1 bag of PBS-T Powder to 500ml of distilled water, and mix well with a magnetic stirrer. The Wash Solution is stable at room temperature for 2 months.

**IMPORTANT NOTES**

1. The wash procedure is critical. Insufficient washing will result in poor precision and falsely elevated RLU values.

2. It is recommended that no more than 32 wells be used for each assay run, if manual pipetting is used, since pipetting of all standards, specimens and controls should be completed within 5 minutes. A full plate of 96 wells may be used if automated pipetting is available.

3. Duplication of all standards and specimens, although not required, is recommended.

**ASSAY PROCEDURE**

1. Secure the desired number of coated wells in the holder. Dispense 25μl of reference standards, specimens, and controls into appropriate wells.

2. Dispense 50μl of Anti-Prog Reagent to each well.

3. Dispense 50μl of Enzyme Conjugate Reagent to each well. Mix gently for 30 seconds.

4. Place on a vortex mixer for 60 minutes at room temperature.

5. At the end of the incubation, remove the incubation mixture by emptying the plate content into a waste container. Rinse and empty the microtiter plate 5 times with washing buffer. Strike the microtiter plate sharply onto absorbent paper or paper towels to remove all residual water droplets. The volume of each well is about 300μl.


7. Incubate at room temperature in the dark for 10 minutes without shaking, then read the RLU values with a Luminolmeter.
CALCULATION OF RESULTS

1. Calculate the mean value from any duplicate reagents. Where appropriate, the mean values should be used for plotting.

2. On linear graph paper plot the RLU (ordinate) for each Reference Standard against the corresponding concentration of PROG in ng/ml (abscissa) and draw a calibration curve through the reference standard points by connecting the plotted points with straight lines.

3. Read the concentration for each control and sample by interpolating on the calibration curve.

4. Computer assisted data reduction will simplify these calculations. If automatic result processing is used, a 4-parameter logistic function curve fitting is recommended.

EXAMPLE OF STANDARD CURVE

A typical standard curve shown below is for the purpose of illustration only, and should never be used instead of the real time calibration curve.

<table>
<thead>
<tr>
<th>P (ng/ml)</th>
<th>RLU</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>12755.2</td>
</tr>
<tr>
<td>0.5</td>
<td>9685.7</td>
</tr>
<tr>
<td>1</td>
<td>7825.3</td>
</tr>
<tr>
<td>2.5</td>
<td>5430.7</td>
</tr>
<tr>
<td>10</td>
<td>2334.8</td>
</tr>
<tr>
<td>30</td>
<td>914.4</td>
</tr>
</tbody>
</table>

EXPECTED VALUES

Each laboratory must establish its own normal ranges based on patient population. The results provided below are based on randomly selected clinical laboratory samples:

progestosterone (ng/ml)

<table>
<thead>
<tr>
<th>Male</th>
<th>Normal Range</th>
</tr>
</thead>
<tbody>
<tr>
<td>Follicular Phase</td>
<td>0.2 ~ 2.4</td>
</tr>
<tr>
<td>Ovulation Phase</td>
<td>0.5 ~ 3.6</td>
</tr>
<tr>
<td>Luteal Phase</td>
<td>6.0 ~ 20.5</td>
</tr>
<tr>
<td>Memopause</td>
<td>0.1 ~ 1.8</td>
</tr>
</tbody>
</table>

PERFORMANCE

A. Sensitivity

The lower detection limit is calculated from the standard curve by identifying the concentration corresponding to the mean RLU of standard diluent (based on 10 replicate analyses) subtract 2 SD. Therefore, the sensitivity of the Autobio Prog CLIA kit is not higher than 0.2 ng/ml.

B. Specificity

No interference was detected with the performance of Autobio Prog CLIA upon addition of massive amounts of the following substances to a human serum pool.

<table>
<thead>
<tr>
<th>Interferents</th>
<th>Concentration</th>
</tr>
</thead>
<tbody>
<tr>
<td>Testosterone</td>
<td>20 ng/ml</td>
</tr>
</tbody>
</table>
Estradiol 1000pg/ml

C. Precision

a. Intra-assay Precision
Intra-assay precision was determined by assaying 20 replicates of each control sera.

<table>
<thead>
<tr>
<th>Serum</th>
<th>Number</th>
<th>Mean</th>
<th>SD</th>
<th>CV (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Low titer</td>
<td>20</td>
<td>2.12</td>
<td>0.07</td>
<td>3.30</td>
</tr>
<tr>
<td>High titer</td>
<td>20</td>
<td>8.31</td>
<td>0.36</td>
<td>4.33</td>
</tr>
</tbody>
</table>

b. Inter-assay Precision
Inter-assay precision was determined by assaying duplicates of each control sera in 10 separate runs.

<table>
<thead>
<tr>
<th>Serum</th>
<th>Number</th>
<th>Mean</th>
<th>SD</th>
<th>CV (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Low titer</td>
<td>10</td>
<td>2.14</td>
<td>0.10</td>
<td>4.67</td>
</tr>
<tr>
<td>High titer</td>
<td>10</td>
<td>8.42</td>
<td>0.41</td>
<td>4.87</td>
</tr>
</tbody>
</table>

E. Accuracy
For samples in the range of 0.2ng/ml to 30ng/ml, the relationship between the Autobio Prog CLIA and the Beckman Access 2® Prog assay, is described by the equation below:

<table>
<thead>
<tr>
<th>Method</th>
<th>No. of Specimens</th>
<th>Linear Equation</th>
<th>Correlation Coefficient</th>
</tr>
</thead>
<tbody>
<tr>
<td>Beckman Access 2®</td>
<td>180</td>
<td>$y = 0.9433x + 0.089$</td>
<td>0.9712</td>
</tr>
</tbody>
</table>

LIMITATIONS
1. For diagnostic purposes, results should be used in conjunction with other data; e.g. symptoms, results of other tests, clinical impressions, etc.
2. If the progesterone results are inconsistent with clinical evidence, additional testing is suggested to confirm the results.
3. Specimens from patients who have received preparations of mouse monoclonal antibodies for diagnosis or therapy may contain human anti-mouse antibodies (HAMA). Such specimens may show either falsely elevated or depressed values when tested with assay kits which employ mouse monoclonal antibodies. Additional information may be required for diagnosis.
4. Heterophilic antibodies in human serum can react with reagent immunoglobulins, interfering with \textit{in vitro} immunoassays. Patients routinely exposed to animals or to animal serum products can be prone to this interference and anomalous values may be observed. Additional information may be required for diagnosis.

QUALITY CONTROL
Good laboratory practice requires that quality control specimens be run with each calibration curve to verify assay performance. To assure proper performance, a statistically significant number of controls should be assayed to establish mean values and acceptable ranges. Controls containing sodium azide should not be used.

SYMBOLS

- **LOT**
- **BATCH CODE**
- **USE BY**
- **MANUFACTURER**
- **CONTAINS SUFFICIENT FOR <n> TESTS**
- **IN VITRO DIAGNOSTIC MEDICAL DEVICE**
- **TEMPERATURE LIMITATION**
- **CATALOGUE NUMBER**
REFERENCES

For order and inquires, please contact
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